

FecalSwab™

Collection, transport & preservation system for enteric pathogens





Copan FecalSwab™ is intended to collect fecal specimens and preserve enteric pathogenic bacteria's viability during transport to the testing laboratory. In the laboratory, FecalSwab™ samples are processed using standard clinical laboratory operating procedures for culture.



FLOQSwabs®



Ensure a quick, capillarity-driven sample uptake and a superior elution of the biological specimen, expanding downstream diagnostic testing capabilities.

Easy to transport



FecalSwab™ simplifies fecal sample collection, transport, and processing. Its shatterproof tubes are a compact and neat alternative to large, bulky transport containers.

Automation compatibility



FecalSwab™ allow to test solid or semi-solid fecal specimens with automatic specimen processors, in space-saving, instrument-ready tubes.

Rectal and stool sampling



FecalSwab™ can be used by medical staff to transfer an adequate quantity of sample from the primary stool collection container or to directly collect a rectal swab sample.

Preservation

FecalSwab™ Performance

Copan FecalSwab™ preserves collected specimens for 48h at room temperature or 72h at refrigerated temperature (2-8°C). In the case of *C. difficile* culture investigation, Copan FecalSwab™ preserves collected specimens up to 24h at room temperature and 48h at refrigerated temperature (2-8°C).

According to the vast scientific literature, FecalSwab™ has been successfully used for:

- Preserve samples at 2-8°C up to 5 days for molecular analysis¹
- Preserve samples at -20°C up to 1 month for molecular analysis²
- Long-term preservation of samples at -80°C for molecular analysis³

FLOQSwabs®

Cut out for everyone

FLOQSwabs® offer **variable sizes, diameters, breaking points and tip shapes to be used in plenty of applications**. This made FLOQSwabs® a well-tolerated alternative to invasive, painful, and costly collection procedures^{7,8}

**Do you have a specific application in mind?
Choose the right FLOQSwabs®!**



Fields of application

Preanalytics made different



Gastrointestinal Diseases^{4,5,6,7}

Regular



Antimicrobial Resistance^{5,8,9}

Regular



Genetics & Microbiome^{2,3}

Regular

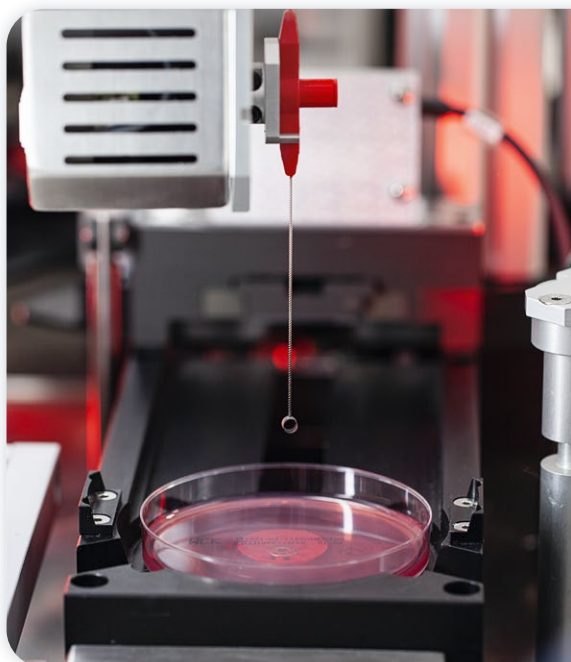
Laboratory

Handling and Processing

In the laboratory, sample processing can be done using manual or automated with Copan WASP¹. Samples collected with FecalSwabTM are suitable for culture of enteric pathogenic bacteria.

Scientific literature reports sample collection transport with FecalSwabTM prior to many downstream diagnostic assays:

- Bacterial culture^{6,8}
- Molecular-based assays^{1,10,11,12}
- Microbiome analysis^{2,3}
- Antigenic assays¹³



WASP®

Walk-Away Specimen Processor™

Copan WASP® is a truly revolutionary instrument for specimen processing for Microbiology. WASP® provides a comprehensive system encompassing all aspects of automated specimen processing, planting and streaking, Gram slide preparation, and enrichment broth inoculation.

Ordering information

Choose between different tube sizes and medium fill volumes, in bulk packs or in combination with either FLOQSwabs® or polyester fiber swabs.

<i>Cat N.</i>	<i>Description</i>	<i>Pack size</i>	<i>Sample*</i>
470CE	FecalSwab™ for manual use, 12x80mm tube filled with 2 ml Modified Cary Blair medium + 1 regular FLOQSwabs® 	500 pieces 10 vipaks of 50 pieces	Stool container, rectal
470CE.A	FecalSwab™ for automation, 12x80 mm tube filled with 2 ml of Modified Cary Blair medium + 1 regular FLOQSwabs® 	500 pieces 10 vipaks of 50 pieces	Stool container, rectal
4E020S.A	FecalSwab™ for automation, 12x80mm rounded bottom tube, filled with 2 ml Modified Cary Blair medium + 1 regular FLOQSwabs® 	500 pieces 10 vipaks of 50 pieces	Stool container, rectal
4E048S	FecalSwab® for manual use, 12x80 mm tube filled with 2 ml of Modified Cary Blair medium + 1 regular FLOQSwabs® with stopper 	500 pieces 10 vipaks of 50 pieces	Stool container, rectal
4U031S	12x80mm tube filled with 2 ml Modified Cary Blair medium + regular FLOQSwabs® 	300 pieces 6 boxes with 50 tubes + 1 resealable pack of 50 stool transfer devices	Stool container

*Suggested table. Please refer to your GLP procedures to choose the most appropriate device for the specific sampling site

Scientific references

All the independent studies we cited in this product focus are listed here.

1. Rojas HF et al (2020) Evaluation of Copan FecalSwab™ preserved stool specimens with the BD MAX™ Enteric Bacterial Panel and the BD MAX™ Extended Enteric Bacterial Panel. *Diagn Microbiol Infect Dis.* 97(4):115055
2. Huey SL et al (2020) Nutrition and the Gut Microbiota in 10- to 18-Month-Old Children Living in Urban Slums of Mumbai, India. *mSphere.* 5(5):e00731-20
3. Biehl LM et al (2019) Usability of rectal swabs for microbiome sampling in a cohort study of hematological and oncological patients. *PLoS One.* 14(4):e0215428
4. Goneau LW et al (2019) Evaluating the preservation and isolation of stool pathogens using the COPAN FecalSwab™ Transport System and Walk-Away Specimen Processor. *Diagn Microbiol Infect Dis.* 94(1):15-21
5. Le Bastard Q et al (2020) Gut microbiome signatures of nursing home residents carrying Enterobacteria producing extended-spectrum β -lactamases. *Antimicrob Resist Infect Control.* 9(1):107
6. Trung NV et al (2017) Non-Typhoidal Salmonella Colonization in Chickens and Humans in the Mekong Delta of Vietnam. *Zoonoses Public Health.* 64(2):94-99
7. Freedman SB et al (2017) Enteropathogen detection in children with diarrhoea, or vomiting, or both, comparing rectal flocced swabs with stool specimens: an outpatient cohort study. *Lancet Gastroenterol Hepatol.* 2(9):662-669
8. Arena F et al (2020) Population structure of KPC carbapenemase-producing *Klebsiella pneumoniae* in a long-term acute-care rehabilitation facility: identification of a new lineage of clonal group 101, associated with local hyperendemicity. *Microb Genom.* 6(1):e000308
9. van Duin E et al (2019) High prevalence of multidrug resistant Enterobacteriaceae among residents of long term care facilities in Amsterdam, the Netherlands. *PLoS One.* 14(9):e0222200
10. Lecronier M et al (2020) Gut microbiota composition alterations are associated with the onset of diabetes in kidney transplant recipients. *PLoS One.* 15(1):e0227373
11. Bhavanam S et al (2020) Differences in Illness Severity among Circulating Norovirus Genotypes in a Large Pediatric Cohort with Acute Gastroenteritis. *Microorganisms.* 8(12):1873
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13. Savolainen R et al (2020) Prospective Evaluation of the mariPOC Test for Detection of *Clostridioides difficile* Glutamate Dehydrogenase and Toxins A/B. *J Clin Microbiol.* 58(4):e01872-19



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